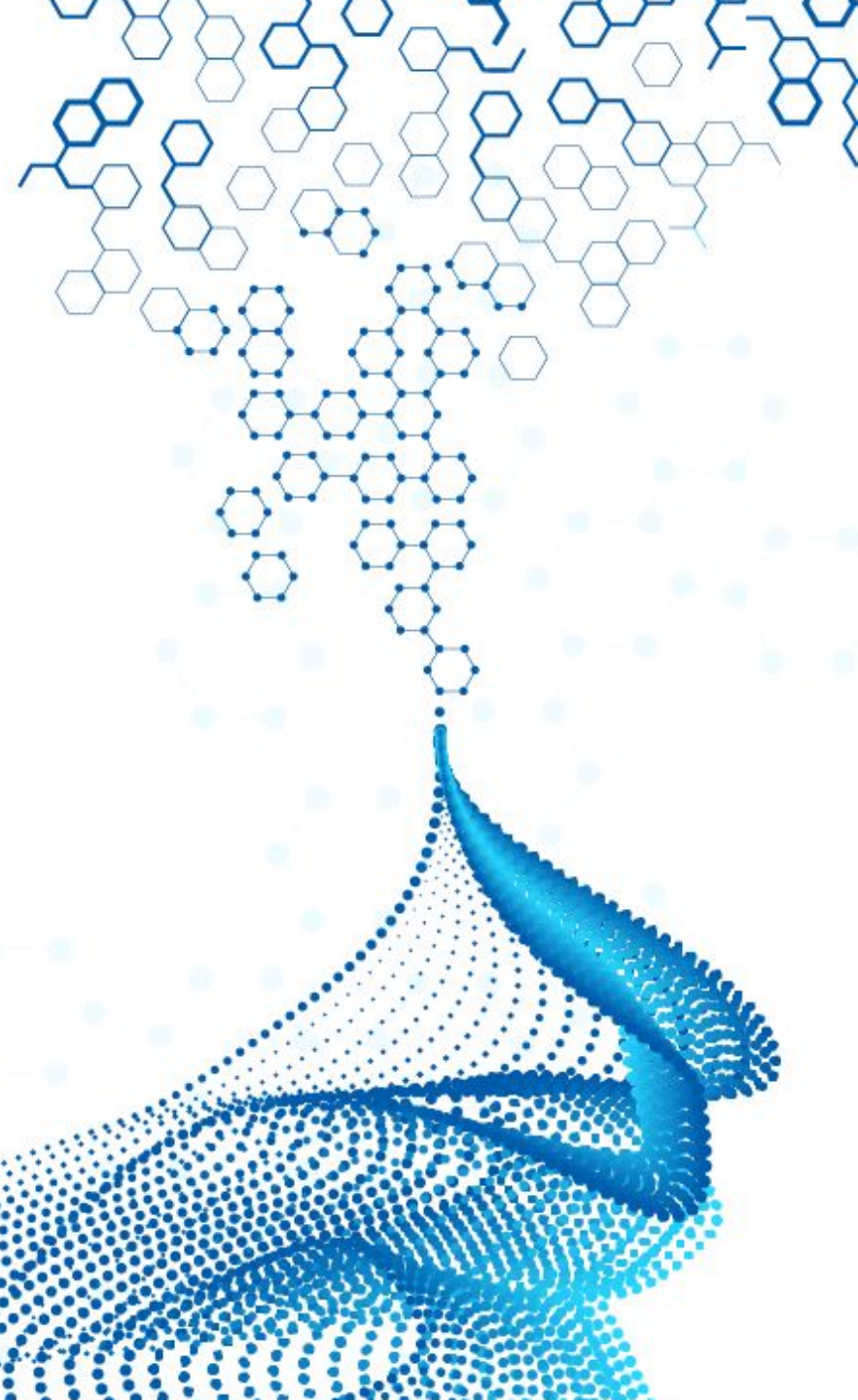




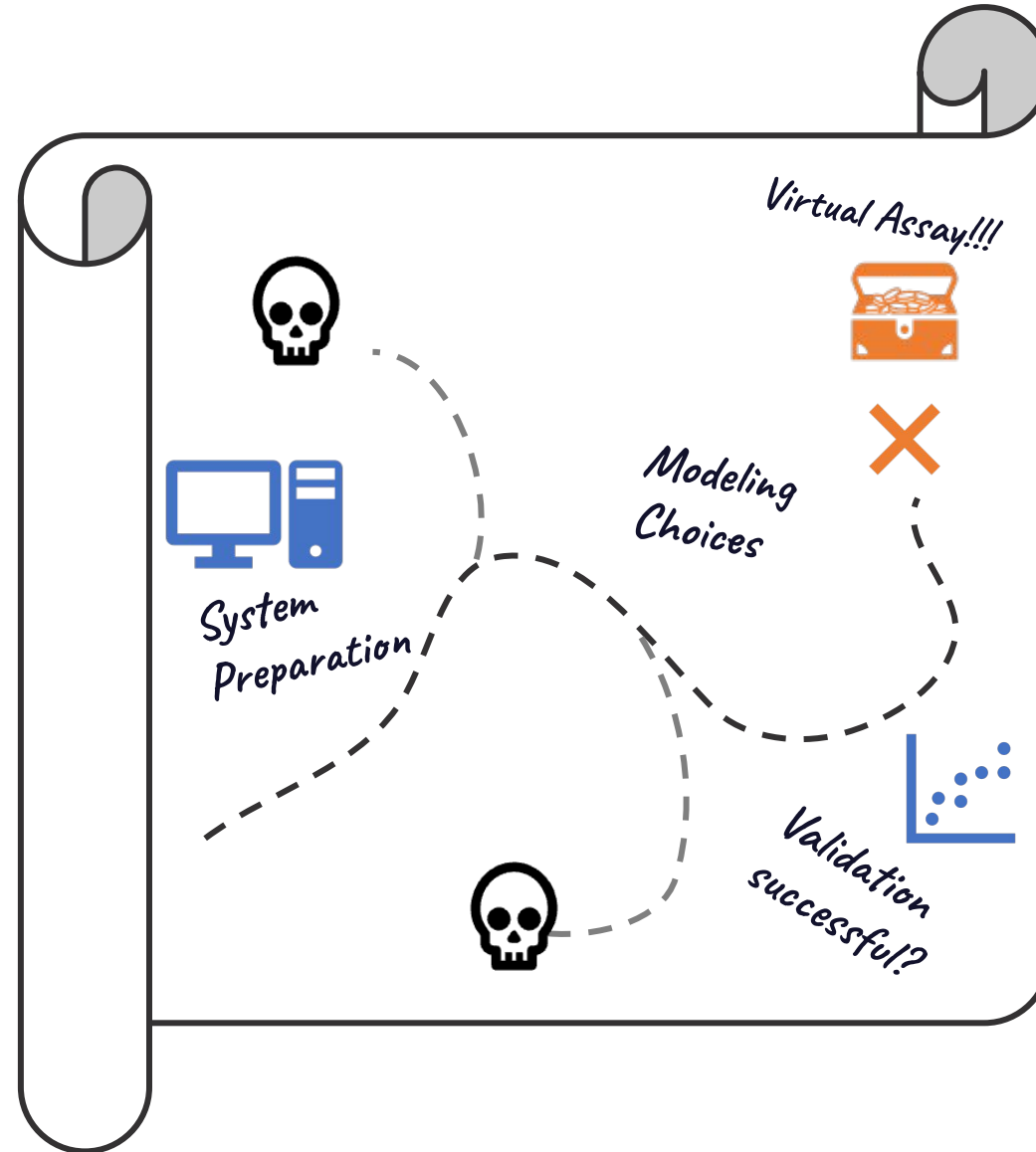
Schrödinger

We need to talk about (Thermo)dynamics

Mila Krämer, Rita Podžuna
LRZ, 2022



Path to Virtual Assay Is Not Necessarily Straightforward



Virtual screening in HitID and LeadOpt

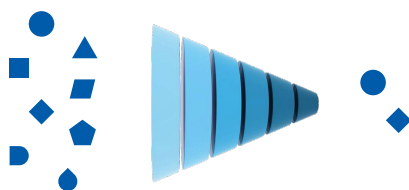
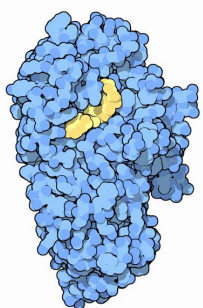
Target selection

Hit identification

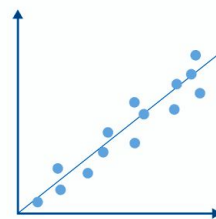
Lead optimisation

Candidate selection

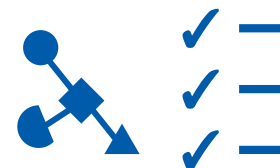
Clinical trials



- Aim of virtual screening is to **filter down large libraries** of diverse compounds
- Requirement is **enrichment** wrt random screening, and to find **diverse hits**
- Additional properties are nice to have but less important at this stage



- Aim of virtual screening is to **rank order congeneric compound ideas**
- Requirement is to **accurately predict binding** and rank similar compounds
- Compounds have to optimally **balance activity and other properties**



Where does Lead Optimization fit in a project?

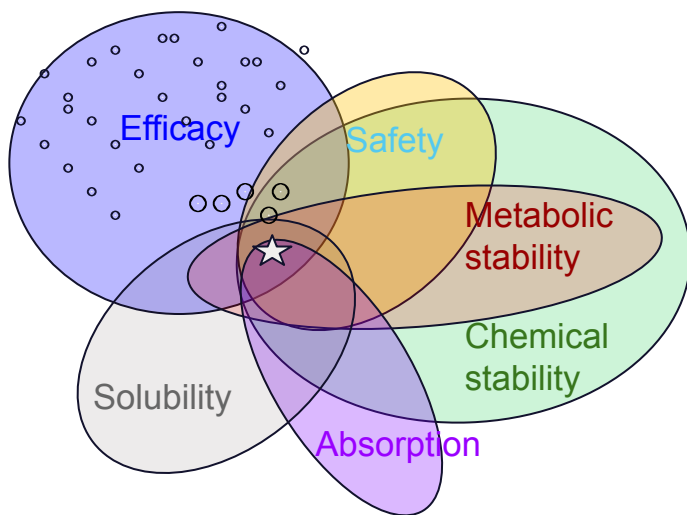
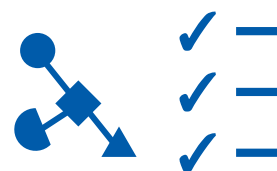
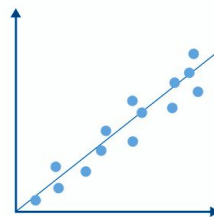
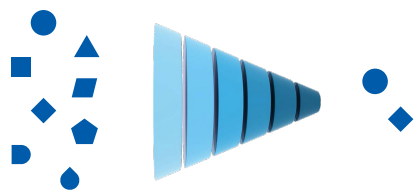
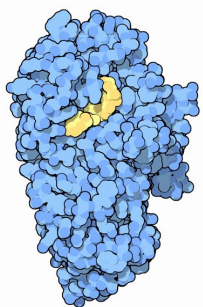
Target selection

Hit identification

Lead optimisation



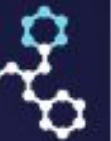



Candidate selection

Clinical trials



- Hit Compounds
- Lead Compounds
- ☆ Drug Candidates

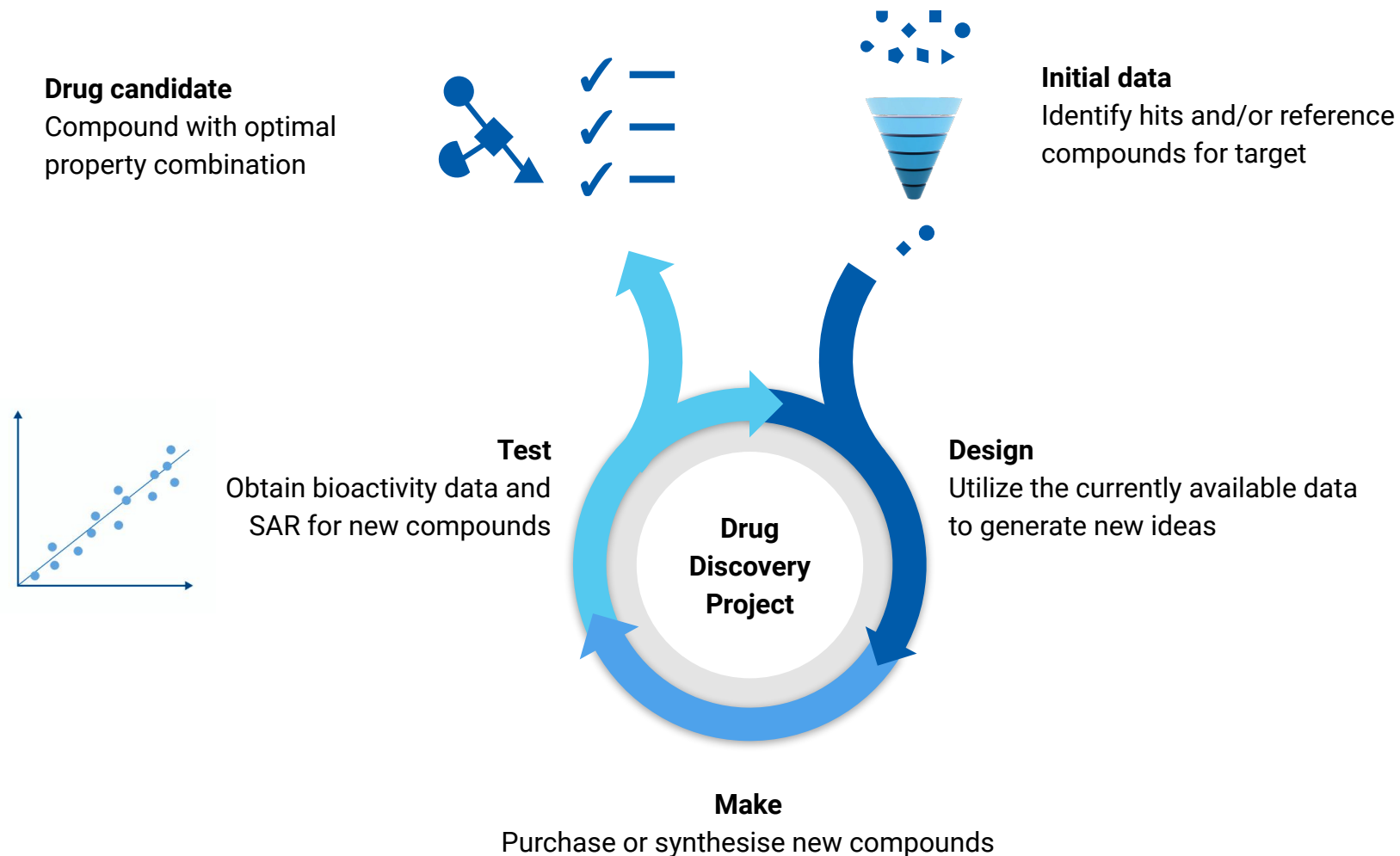
Designing drugs is an extremely hard multi-dimensional optimization problem

Need to identify a molecule that balances a large number of anti-correlated properties:						
Potency	✓	✗	✓	✗	✓	✓
Selectivity	✗	✓	✓	✓	✗	✓
Solubility	✗	✗	✗	✓	✓	✗
Bioavailability	✗	✗	✗	✗	✗	✗
Clearance / half-life	✗	✗	✗	✗	✗	✗
Permeability	✗	✗	✗	✗	✗	✗
Drug-drug interactions	✗	✗	✗	✗	✗	✗
Synthesizability	✗	✗	✗	✗	✗	✗
Toxicity	✗	✗	✗	✗	✗	✗

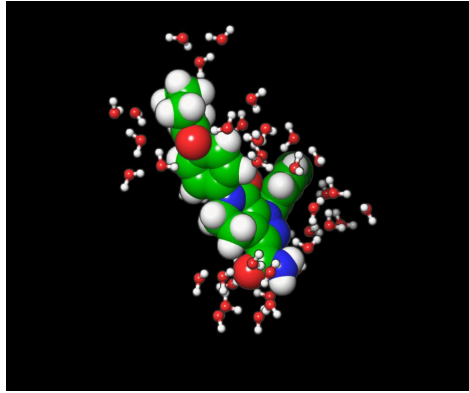


* Based on average, industry-wide success rates

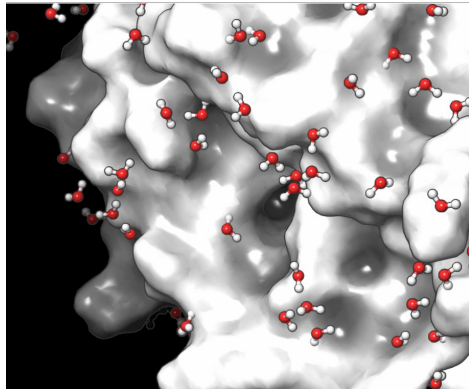
Lead Optimization is Complex, Slow and Expensive



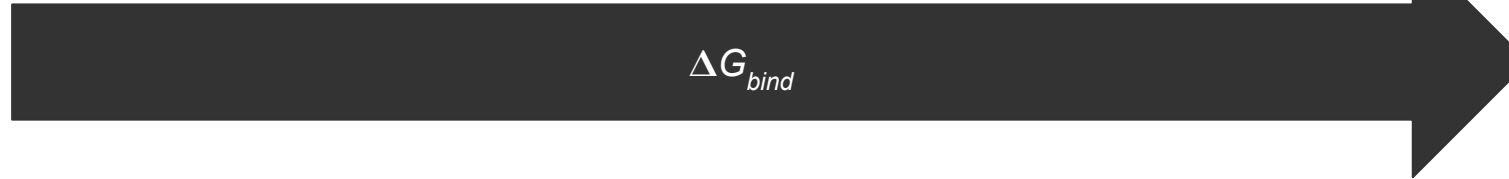
One way to look at protein-ligand binding



*Flexible molecule
in water*

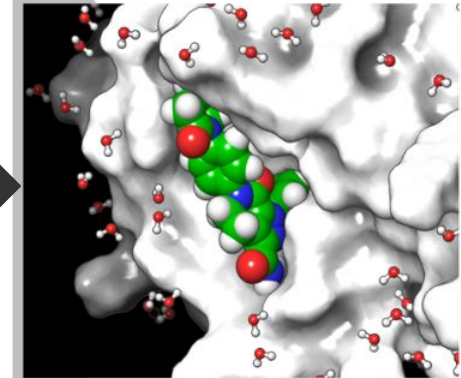


*Flexible protein
in water*



$$\Delta G_{bind}$$

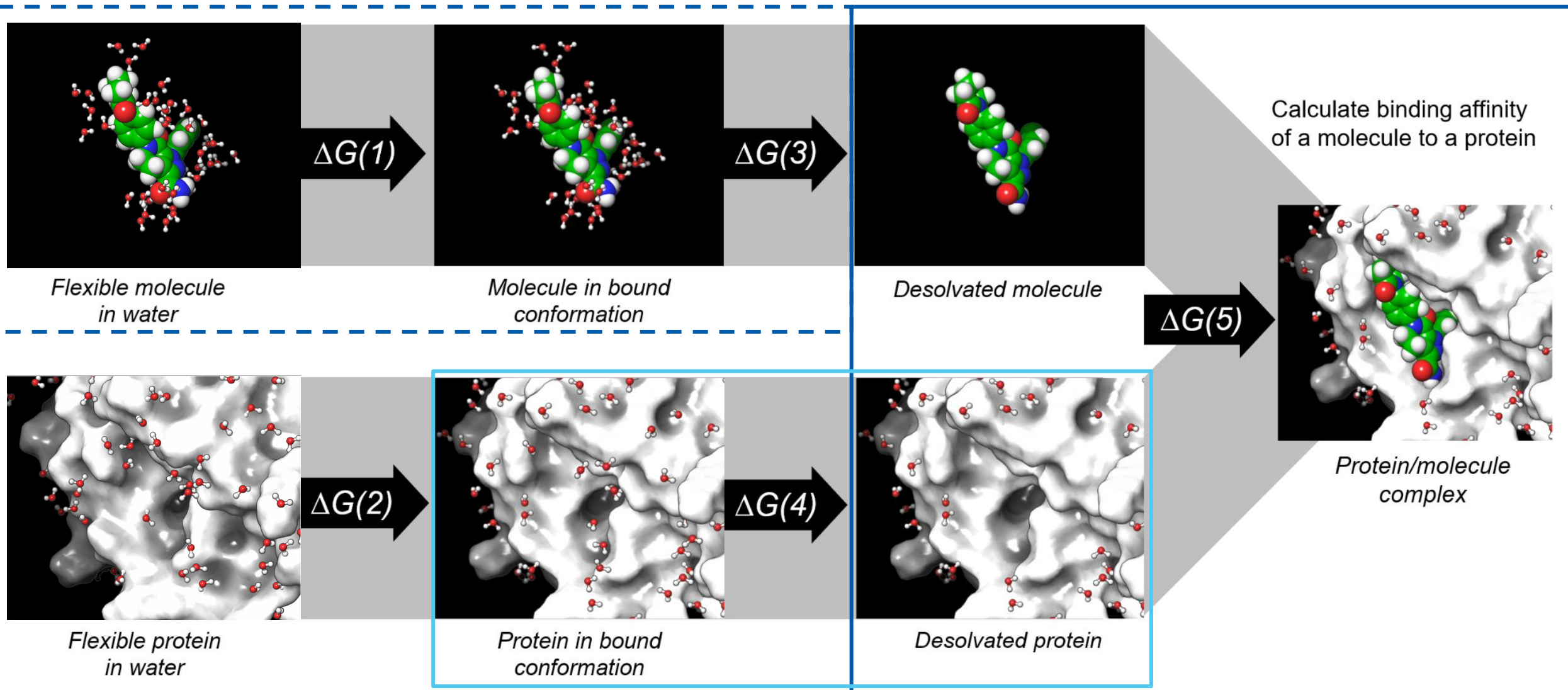
Calculate binding affinity
of a molecule to a protein



*Protein/molecule
complex*

One way to look at protein-ligand binding

Docking (Glide)

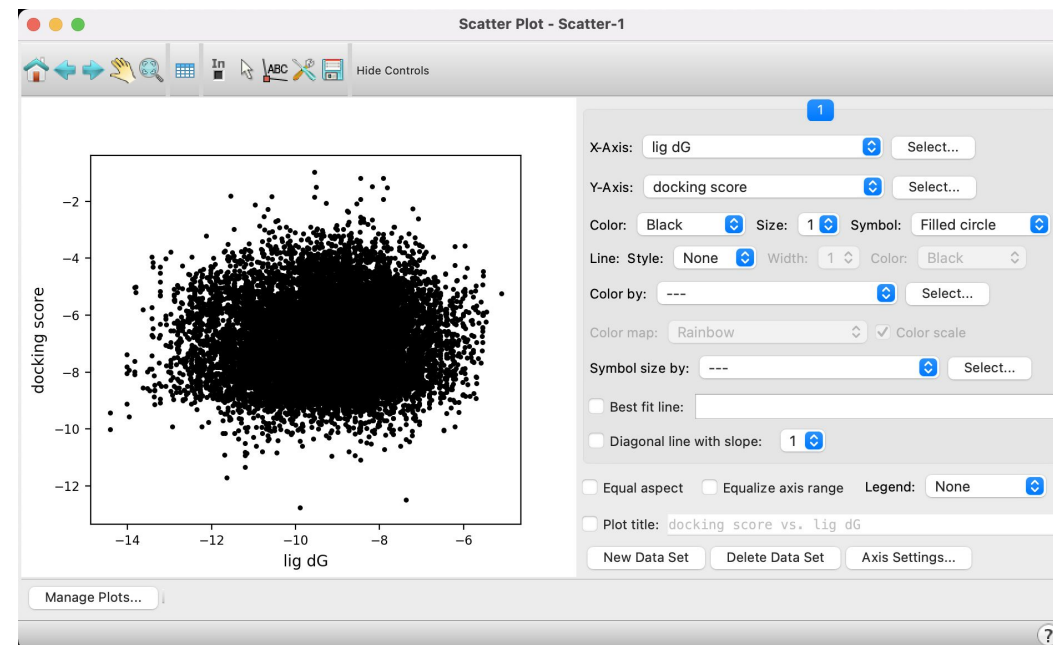


WaterMap

$$\Delta G_{bind} = \Delta G(1) + \Delta G(2) + \Delta G(3) + \Delta G(4) + \Delta G(5)$$

Correlation of Docking Results to Binding Affinity

- In many compound sets, there is only a very weak correlation between docking score and experimental binding affinity or none at all.
- Generally docking score can not be used to distinguish between less and more active compounds



⇒ Reminder: the docking score is parametrized to efficiently distinguish binders from non-binders, not as a proxy for binding affinity

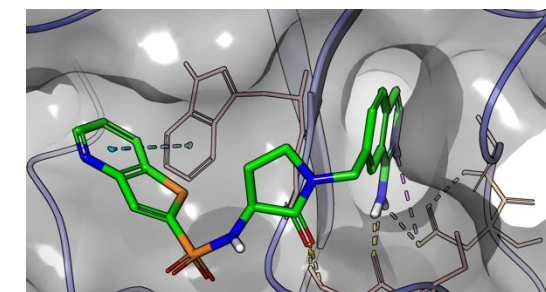
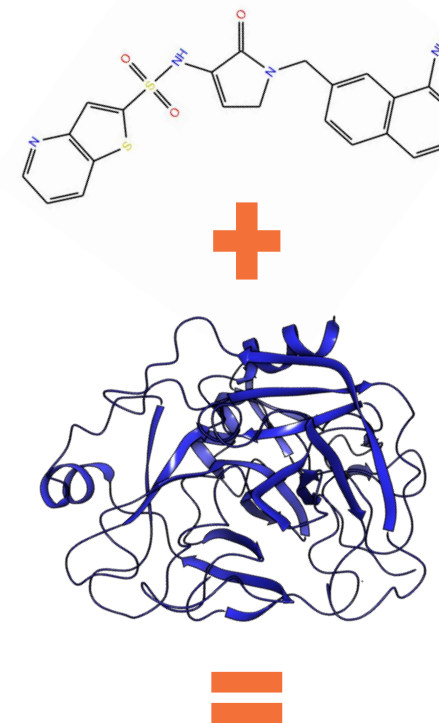
Binding Affinity Prediction from Static Structures

Limitations:

- 3D structure(s):
 - Homology or X-ray (but which co-crystal ligand?)
 - Flexibility of the receptor and ligand often marginally included (sometimes via multiple structures)
 - Experimental conditions might not be reflected by the structure
- Computational model:
 - Implicit solvent models: continuum dielectric models do not reflect the complex effects of microsolvation
 - Force fields: need to reproduce the energy gain upon complex formation but also the relative energy gain upon solvation

Results:

- Out-of-the-box correlation with experimental data can be poor
- Experience plays a crucial role in correcting these limitations



GlideScore = - 7.42
MMGBSA Score = - 9.01

Free Energy Perturbation - Calculating Relative Binding Energies

The diagram illustrates the calculation of relative binding energies using Free Energy Perturbation. It shows the binding of ligand A and ligand B to a protein, and the corresponding free energy changes for the perturbation from A to B in both the free state and the bound state.

Top row: Protein + Ligand A $\xrightarrow{\Delta G_{\text{Bind}}^A}$ Protein-A complex

Bottom row: Protein + Ligand B $\xrightarrow{\Delta G_{\text{Bind}}^B}$ Protein-B complex

Left vertical arrow: Protein + Ligand A $\xrightarrow{\Delta G_{\text{Alc-Solv}}^{A \rightarrow B}}$ Protein + Ligand B

Right vertical arrow: Protein-A complex $\xrightarrow{\Delta G_{\text{Alc-Cmplx}}^{A \rightarrow B}}$ Protein-B complex

$$\Delta\Delta G_{\text{Bind}}^{A \rightarrow B} = \underbrace{\Delta G_{\text{Bind}}^B - \Delta G_{\text{Bind}}^A}_{\text{hard but interesting}} = \underbrace{\Delta G_{\text{Alc-Cmplx}}^{A \rightarrow B} - \Delta G_{\text{Alc-Solv}}^{A \rightarrow B}}_{\text{simpler to calculate now}}$$

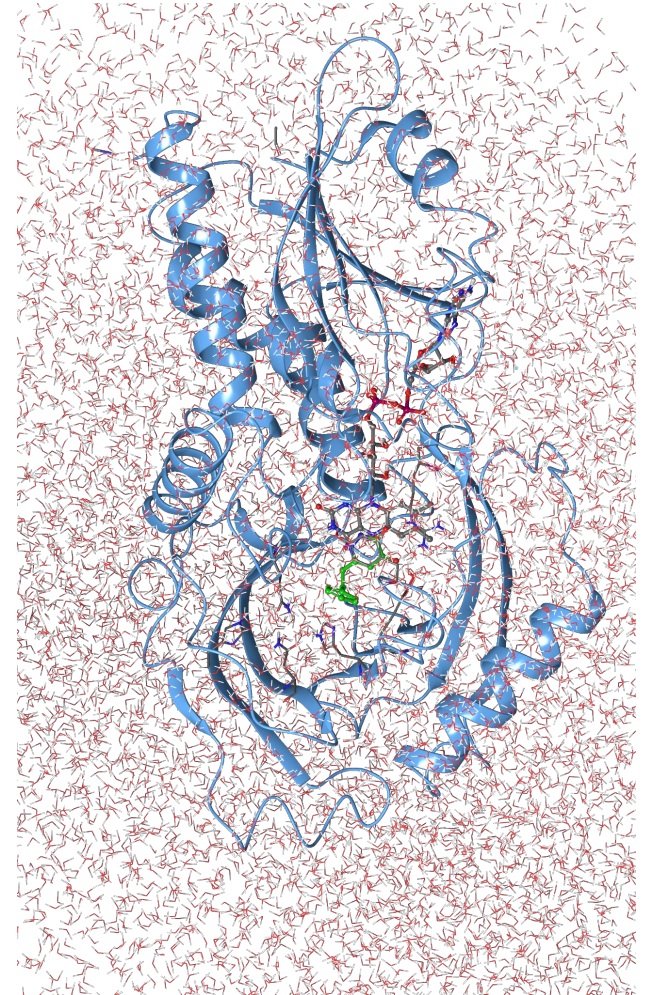
Understanding Hydration

using WaterMap



Reminder: Why is Water Important?

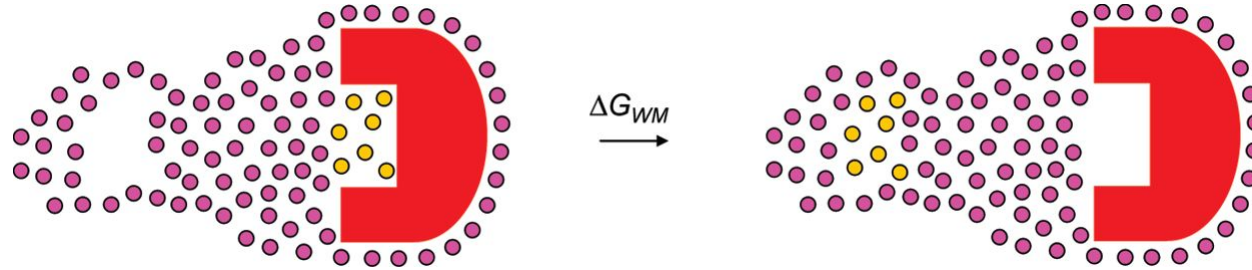
- Water is everywhere in biology
- “Empty” binding sites are mostly filled with water
- Ligands must displace that water to bind



⇒ Water energetics can drive potency, but can't be obtained from static structures

What do ΔG , ΔH and $-T\Delta S$ correspond to?

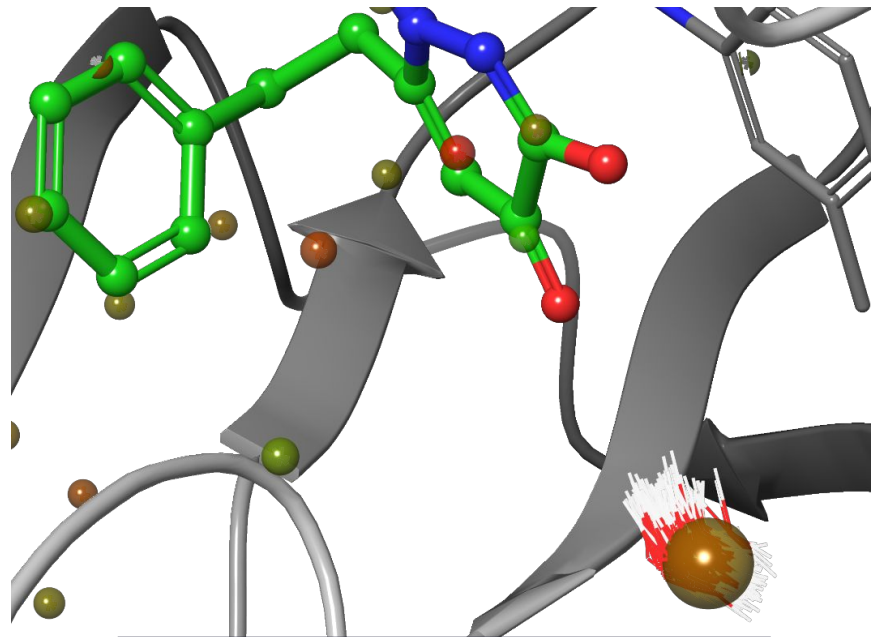
- The values calculated by WaterMap correspond to the average excess enthalpy, entropy and free-energy that a water molecule, located at the hydration site, would possess relative to bulk water



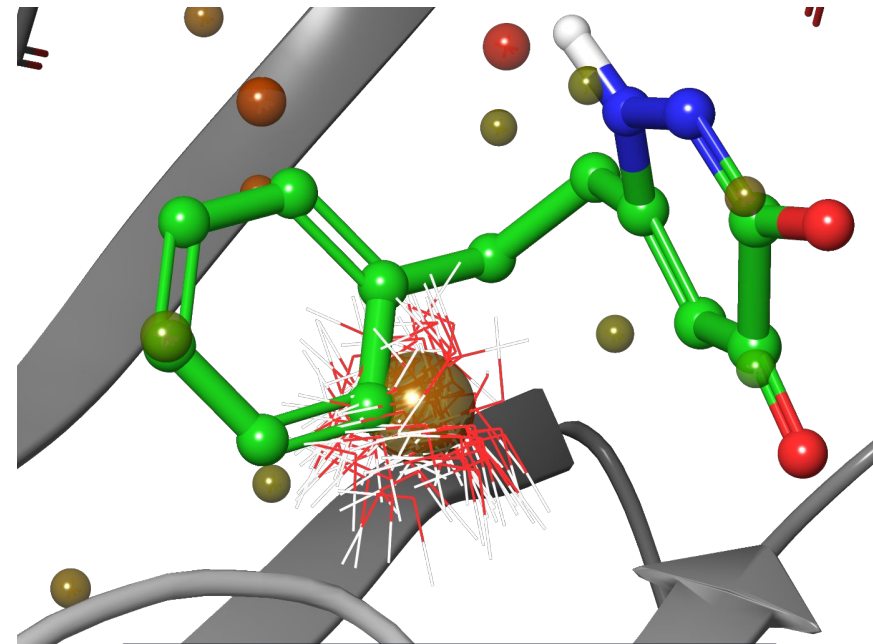
- This means that:
 - A hydration site with a **negative** ΔH -value is making **stronger** interactions with the surrounding protein than it would with surrounding water molecules in solution e.g. near a charged group
 - A hydration site with a **positive** ΔH -value is making **weaker** interactions with the surrounding protein than it would with surrounding water molecules in solution e.g. near a hydrophobic residue

What do ΔG , ΔH and $-T\Delta S$ correspond to?

- The offensive mathematics is just quantifying the 'randomness' of the water molecules at each hydration site



Highly ordered water molecules
Unfavourable entropy
 $-T\Delta S=5.29$ kcal/mol

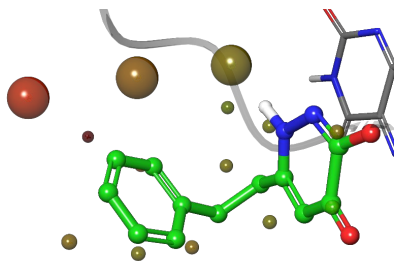


Disordered water molecules
Favourable entropy
 $-T\Delta S=0.71$ kcal/mol

M Analyzing WaterMap Results

show waters occupying hydration sites

select sites in workspace to highlight them and select corresponding rows in table



filter sites by properties

WaterMap - Examine Results

Load results: Adjust the view when analyzing

Display: Receptor Ligand H-bonds Markers Shape by entry Dipoles
 Waters Ligand surface Water density Cavity map Free energy density

Site label: None

Color sites by: ΔH and $-\Delta S$ Color scale: Relative Absolute Show color in table

Site	Occupancy	Overlap	ΔH	$-\Delta S$	ΔG	#HB(WW)	#HB(PW)	#HB(LW)
47	0.29	0.00	-0.52	0.59	0.07	2.70	0.00	0.00
45	0.31	0.00	1.05	0.70	1.75	2.48	0.00	0.00
42	0.35	0.00	0.19	0.71	0.90	2.43	0.00	0.00
46	0.31	1.00	2.56	0.71	3.27	2.19	0.00	0.00
43	0.34	0.00	0.19	0.75	0.94	2.44	0.02	0.00

Pick to select sites Show Only Selected Rows

Show only water sites with:

Enthalpy (ΔH) kcal/mol Invert range

Entropy ($-\Delta S$) kcal/mol Invert range

Free energy (ΔG) kcal/mol Invert range

Overlap factor Invert range

Distance Å of entry.id 13

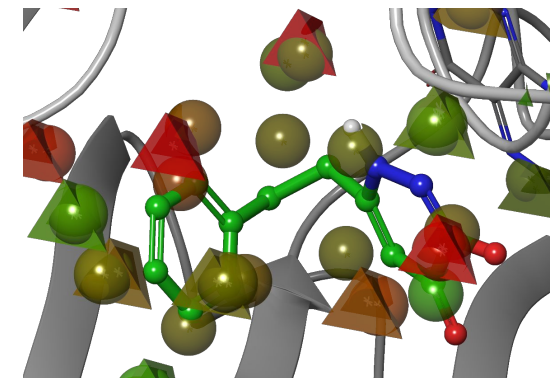
Score Ligands Create properties for selected sites Score only selected sites

Perform WM/MM Scoring...

import raw MD simulation data (rarely necessary)

reset panel to return to blank slate

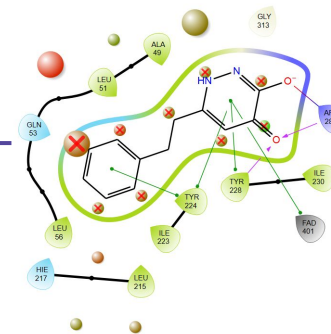
useful for comparing multiple WaterMaps:



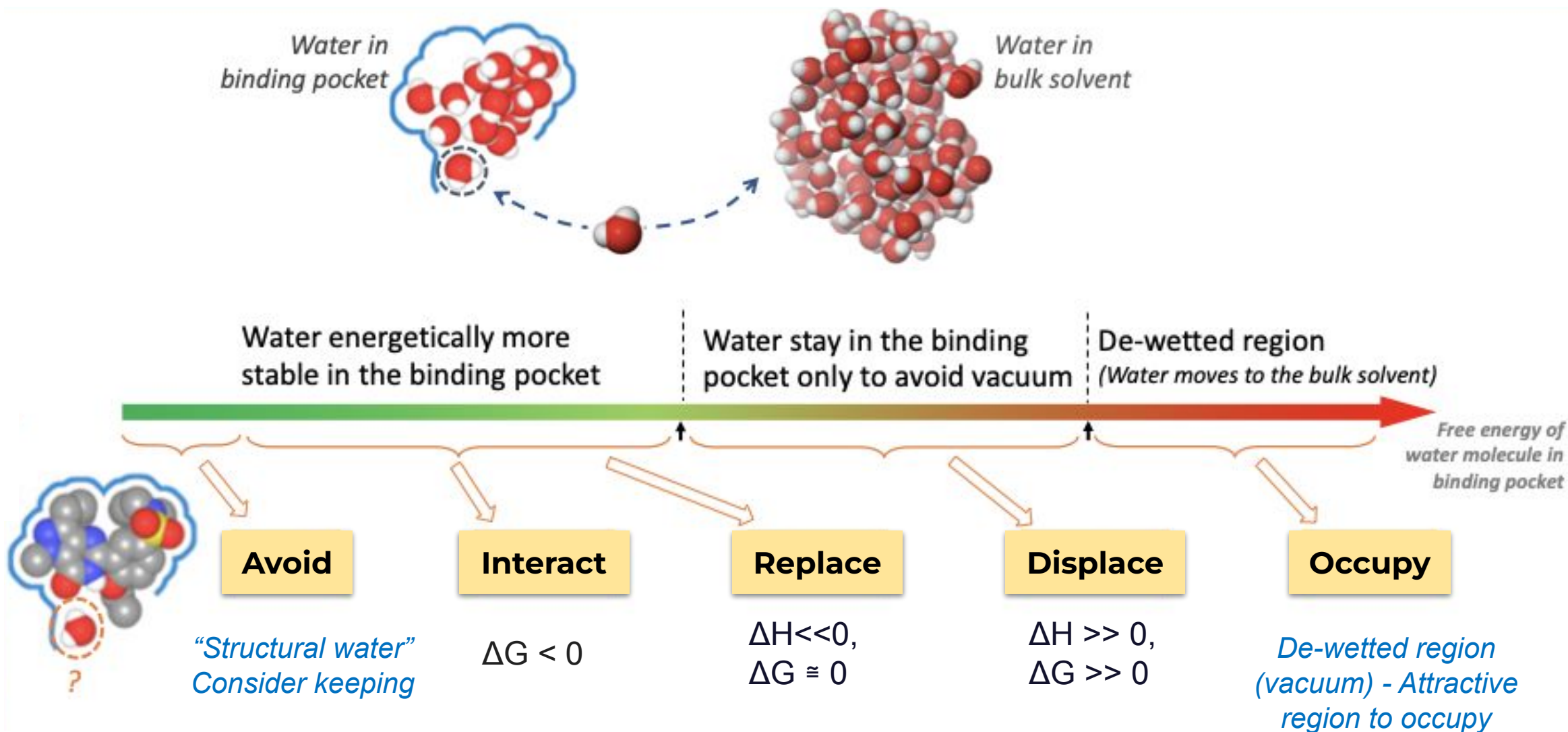
Comparing apo (spheres) vs holo (pyramids) WaterMap highlights waters displaced by ligand

visualize water density, cavities and continuous WaterMap

2D view



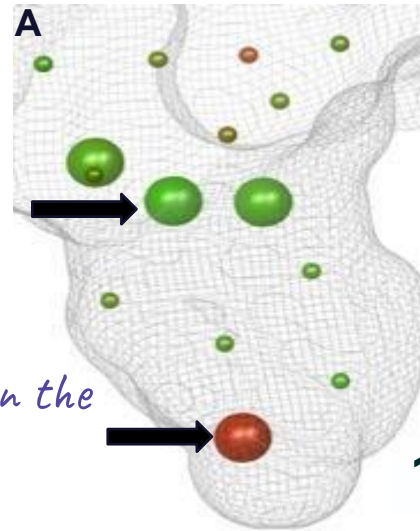
How can Understanding Hydration Guide Strategy?



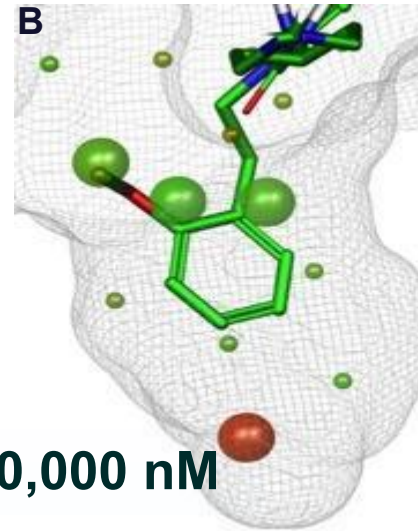
Mapping out where water molecules are can guide and explain SAR

3 stable waters with favorable enthalpy

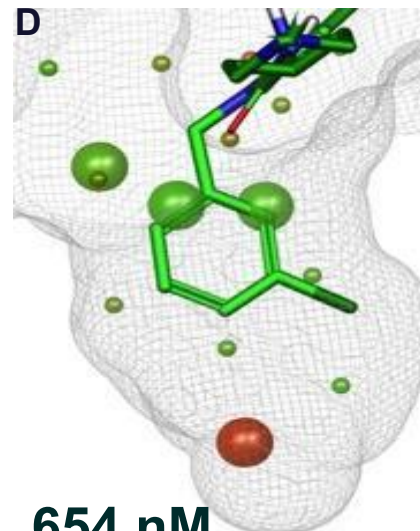
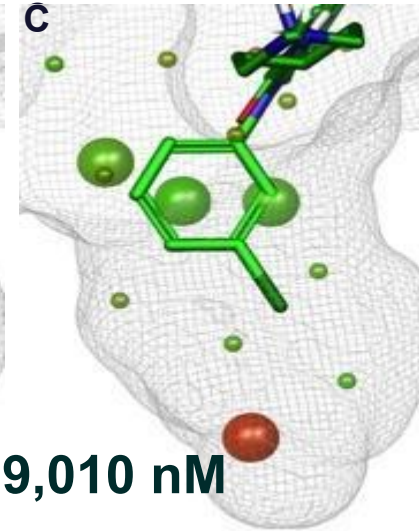
most unstable water in the binding site



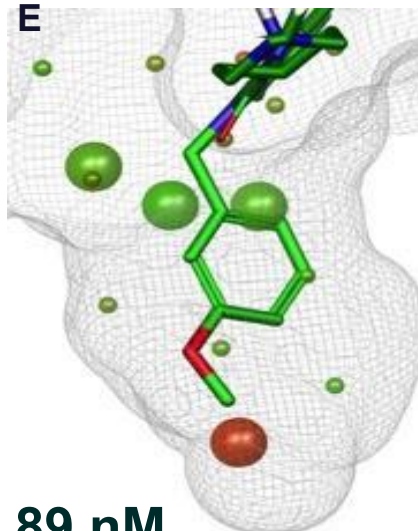
10,000 nM



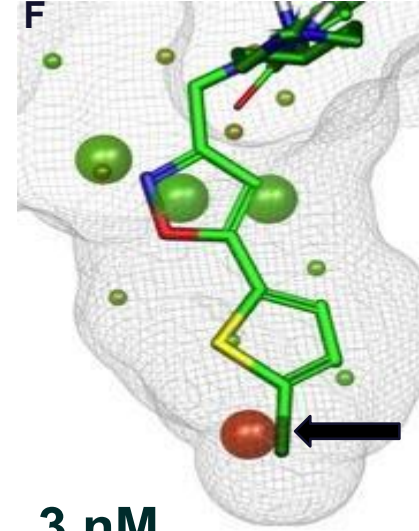
9,010 nM



654 nM



89 nM



3 nM

full displacement of unstable water corresponds to tightest binder



Schrödinger

Thank You!

mila.kraemer@schrodinger.com